

2. Fieldwork and analytical programme

2.1. Survey execution and sample collection

The survey was conducted from 29 July – 3 August 2010, the sampling log is given in appendix 1. The survey vessel was the *Svetlomor 2*, operated by CBARS. The following personnel participated in the survey:

- James McNee, AmC Senior Consultant
- Ilgar Guliyev, AmC Senior Scientist
- Mustafa Gafarov, AmC Scientist
- Irada Israfilova, AmC Scientist
- Konul Guliyeva, AmC Scientist
- Faruhk Fursi, AmC Technician
- Kamran Ahmedov, AmC Technician
- Andrey Fomin, AmC Technician
- Valeh Karimov MENR Scientist
- Hikmet Zeynalov SOCAR Scientist

2.2. Sampling & Analytical Programme

Details of the analytical methods are provided in Appendix 2 of this report. The analytical work was conducted at AmC Caspian Environmental Laboratory. All sampling and laboratory work was carried out in accordance with AmC's quality system.

A standard sediment survey was carried out, using a double 0.1 m² Van Veen grab to collect samples from the top 10-15 cm of sediment. Two replicate samples were collected at each station for physical (particle size) and chemical analysis and three replicate samples were collected for biological analysis (see table 2.1). Replicate independence was assured by processing one biology and one physical/chemistry sample from two of the three double grabs and a single biology sample from the third double grab.

Biological samples were washed on a 0.5 mm sieve, and all material retained was preserved in 4% formaldehyde solution. Subsamples for physical and chemical analysis were placed in appropriate labelled containers and stored in a freezer on the *Svetlomor 2*, then transferred to freezer storage at AmC's Caspian Environmental Laboratory at the end of the survey. Sample station coordinates and the number of samples taken for each analysis is detailed in table 2.1.

The bioassays was conducted following the Caspian Specific Ecotoxicology Protocol (CSEP). The sediment bioassay procedure uses the Caspian amphipod *Pontogammarus maeoticus*, which is exposed directly to the sediment for 96 hours. Tests were conducted in 1-litre glass vessels, each containing approximately 250 grammes of sediment, 700 ml of filtered Caspian seawater, and 10 adult amphipods. Once preliminary data on contamination levels were available, eight stations were selected for assessment of the toxicity of the sediment by exposing animals to the sediment in laboratory conditions (bioassay). These stations were selected to give a full range of contamination, and maximum possible spatial coverage. Two field replicate samples from each station were tested and the laboratory tests were conducted in duplicate (ie four tests conducted for each station).

Water samples were collected above and below the thermocline at stations 1, 8, 9 & 15 using a 12l Niskin Sampler, the water analysis plan is given in table 2.2.

Table 2.1 Co-ordinates of Seabed Stations & Sampling Plan

Station	Easting	Northing	Depth	Physical and Chemical Analysis						Biological Analysis		Water
				THC	PAH	Phenols	HM	Total Ba	PSA	Bioassay	Taxonomy, Abundance and Biomass	
CH1001	520428	4439465	128	2	2	2	2	2	2	2	3	2
CH1002	520385	4439310	131	2	2	2	2	2	2	2	3	
CH1007	520428	4438715	117	2	2	2	2	2	2	2	3	
CH1008	520428	4438465	117	2	2	2	2	2	2	2	3	2
CH1009	519853	4438988	167	2	2	2	2	2	2	2	3	2
CH1015	521049	4438997	128	2	2	2	2	2	2	2	3	2
CH1016	521277	4438116	112	2	2	2	2	2	2		3	
CH1025	519579	4439814	153	2	2	2	2	2	2		3	
CH1033	521277	4439814	143	2	2	2	2	2	2	2	3	
CH1034	520601	4439199	126	2	2	2	2	2	2	2	3	
CH1035	520645	4438840	117	2	2	2	2	2	2	2	3	
CH1036	520226	4438735	130	2	2	2	2	2	2	2	3	
CH1037	520160	4439134	113	2	2	2	2	2	2	2	3	
CH1038	520725	4439375	130	2	2	2	2	2	2		3	
CH1039	520962	4439269	130	2	2	2	2	2	2		3	
CH1040	520861	4438715	117	2				2				
CH1041	520678	4438532	115	2				2				
CH1042	520178	4438532	127	2				2				
CH1043	519943	4438663	147	2				2				
CH1044	519995	4439215	156	2				2				
CH1045	520178	4439398	130	2				2				
CH1046	520958	4439495	135	2	2	2	2	2	2	2	3	
CH1047	520958	4438401	121	2	2	2	2	2	2	2	3	
CH1048	519898	4438435	156	2	2	2	2	2	2	2	3	
CH1049	519898	4439495	142	2	2	2	2	2	2	2	3	
CH1050	520510	4440205	142	2	2	2	2	2	2		3	
CH1051	521428	4438965	128	2				2				
CH1052	520428	4437965	118	2	2	2	2	2	2		3	
CH1053	519537	4438956	173	2	2	2	2	2	2		3	
CH1054	519593	4438130	171	2	2	2	2	2	2	2	3	
CH1055	521489	4440026	146	2	2	2	2	2	2		3	
CH1056	521842	4440379	153	2	2	2	2	2	2	2	3	
CH1057	522549	4441086	177	2	2	2	2	2	2		3	

- THC by GC-FID, 2-6 ring PAH by GC-MS
- Phenols measured using spectrophotometry (Merck Spectroquant method)
- HM = Heavy metal analysis: All metals except Hg analysed by ICP-OES, Hg analysed by CVAFS
- PSA = Particle size analysis: Grain size distribution, total organic matter (TOM), carbonate (CO₃)

Table 2.2 Water Sampling Analysis Plan

Determinand	Method	Method of sampling and preservation for transport to laboratory
Temperature	pH meter	N/A – analyses conducted immediately
Salinity	Multimeter	
Dissolved oxygen	DO meter	
Turbidity	Turbidimeter	
pH	pH meter	
Total suspended solids (TSS)	EN 872	Water is filtered through pre-weighed membrane filter. Filter is placed in labelled plastic Petri dish, dish is sealed and frozen.
Total nitrogen, total phosphorus, nitrate, nitrite, ammonia, phosphate, silicate, COD & BOD	Grasshoff <i>et al</i>	Water is poured into two (duplicate) sterile 1-litre plastic bottles, then frozen. 2 litres of water is sufficient for all nutrient analyses.
THC	GC-FID	Water is poured into 2 x 1litre, DCM-cleaned, amber glass bottle, then refrigerated
PAH	GC-MS	
Phenols	Merck Spectroquant	
Metals (Fe, Cd, Co, Cu, Ni, Pb & Zn)	ICP-MS	Water is poured from Niskin bottle into 0.5-L sterile plastic bottles, then refrigerated